

StabyExpress™ kit Manual for GST fusion

(v1.0) for GE-SEGST-.... kits



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Content and storage :

The StabyExpress™ expression kit is shipped on **dry ice**.

Storage: -80°C

Two different types of StabyExpress™ kits are available: one containing electrocompetent cells (in non self-standing tubes), and the other type containing chemically-competent cells (in self standing tubes).



Each kit contains one box with the following items:

Name	Concentration / remarks	Amount (SEGST0505, SEGST0707)	Amount (SEGST1010, SEGST1212)	Amount (SET70020, SET70022)	Amount (SET71111, SET71313)
pStabyGST1.2 DNA	0.1µg/µl	1 tube of 50µl	1 tube of 50µl	/	/
CYS21 strain (for cloning) Pink cap	Competent cells	5 tubes	10 tubes	/	10 tubes
SE1 strain (for expression) Blue cap	Competent cells	5 tubes	10 tubes	20 tubes	10 tubes
Staby reverse primer Red cap	0.1µg/µl in water 5'-CCAACTCAGCTTCCTTCG-3'	1 tube of 20µl	1 tube of 20µl	/	/
Staby forward primer Red cap	0.1µg/µl in water 5'-GCGTCCGGCGTAGAGGATC-3'	1 tube of 20µl	1 tube of 20µl	/	/
Regeneration medium White cap	2% Tryptone 0.5% Yeast extract 0.05% NaCl 2.5mM KCl 10mM MgCl ₂	5 tubes of 1.5ml	10 tubes of 1.5 ml	20 tubes of 1.5 ml	10 tubes of 1.5ml
Manual		1	1	1	1

The genotype of the CYS21 strain is: F⁻, Cm^R, *mcrA*, *endA1*, $\Delta(mrr-hsdRMS-mcrBC)$ (restriction-, modification-), $\Phi 80lacZ\Delta M15$, $\Delta lacX74$, *recA1*, $\Delta(ara, leu)7697$, *araD139*, *galU*, *galK*, *nupG*, *rpsL*, *ccdB⁺*.

The genotype of the SE1 strain is: derivatives from *E. coli* B strain, F⁻, Cm^R, *ompT*, *lon*, *hsdS_B* (restriction-, modification-), *gal*, *dcm*, DE3 (*lacI*, T7 polymerase under the control of the PlacUV5 promoter), *ccdB⁺*.

Material Safety Data Sheet:

Product and company identification:

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Fax: +32.71.37.60.57
e-mail: delphigenetics@delphigenetics.com

Hazards identification

No specific hazard concerning the products of the StabyExpress T7 kit.

First aid measures

- Inhalation: If one of the products of the StabyExpress T7 kit is inhaled, remove to fresh air. If not breathing, give artificial respiration. If breathing is difficult, give oxygen. Get medical attention.
- Ingestion: Do not induce vomiting unless directed to do so by medical personnel. Never give anything by mouth to an unconscious person. If large quantities of the products of the StabyExpress T7 kit are swallowed, call a physician immediately.
- Skin contact: In case of contact, immediately flush skin with plenty of water. Remove contaminated clothing and shoes. Wash clothing before reuse. Thoroughly clean shoes before reuse. Get medical attention.
- Eye contact: In case of contact with one of the products of the StabyExpress T7 kit, immediately flush eyes with plenty of water for at least 15 minutes. Get medical attention.

Fire-fighting measures

Use foam or all purpose dry chemicals to extinguish. Fire fighters should wear positive self-contained breathing apparatus and full turnout gear.

Accidental release measures

Immediately contact emergency personnel. Use suitable protective equipment (see below exposure controls and personal protection). For small spills add absorbent, scoop up material and place in a sealed, liquid-proof container for disposal. For large spills dike spilled material or otherwise contain material to ensure runoff does not reach a waterway. Place spilled material in an appropriate container for disposal. Minimize contact of spilled material with soils to prevent runoff to surface waterways.

Handling and storing

Keep the container tightly closed, in a cool and well-ventilated area.

Personal protection

The occupational exposure limits were not determined. Protect your skin and body using uniform or laboratory coat, chemical resistant, impervious gloves. Use safety glasses, face shield or other full-face protection if potential exists for direct exposure to aerosols or splashes.

Disposal consideration

Waste must be disposed of in accordance with federal, state and local environmental control regulations.

N.B.: Final determination of suitability of any material is the sole responsibility of the user. All materials may present unknown hazards and should be used with caution. To the best of our knowledge, the information contained herein is accurate. However, neither Delphi Genetics SA nor any of its subsidiaries assumes any liability whatsoever for the accuracy or completeness of the information contained herein.

Licenses

The StabyExpress™ expression kit is covered by worldwide patents (US7183097, US7595186, US7595185 and other patents pending). The kit is sold under a license from the Université Libre de Bruxelles (Belgium). **The kit is sold for research purpose only.** A license from Delphi Genetics SA is required for any commercial use.

(Please, contact Delphi Genetics at delphigenetics@delphigenetics.com)

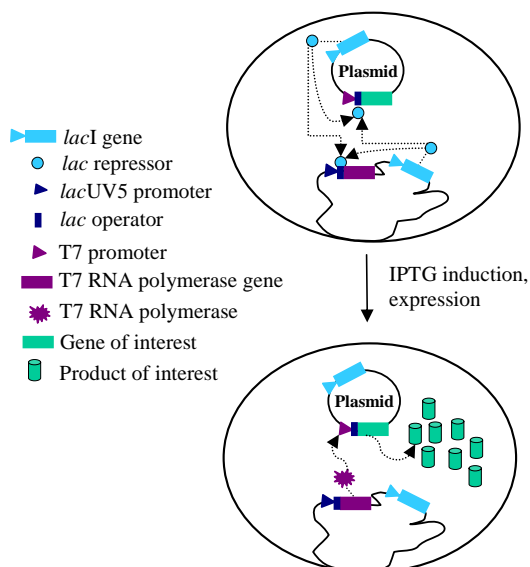
T7 expression kit is based on technology developed at Brookhaven National Laboratory under contract with the U.S. Department of Energy and is the subject of patent applications assigned to Brookhaven Science Associate (BSA) in the United States of America.

User Guide

Overview of the T7 expression system

The T7 expression system is based on the use of the T7 bacteriophage promoter and RNA polymerase. The T7 RNA polymerase is useful for synthesizing large amounts of RNA selectively: the T7 RNA polymerase only recognizes the T7 promoter and not the *E. coli* promoters. Conversely, the *E. coli* RNA polymerase does not recognize the T7 promoter (see below). The T7 RNA polymerase is able to transcribe genes five times faster than the *E. coli* RNA polymerase (Chamberlin and Ring, 1973; Golomb and Chamberlin, 1974). The gene encoding the T7 RNA polymerase is inserted into the chromosome of the expression bacteria (SE1, figure 1). The expression of this gene is under the control of the *lacUV5* promoter and therefore is basically controlled by the same mechanisms as the *lac* operon. Thus, the expression of the T7 RNA polymerase is repressed by the binding of the *lac* repressor (encoded by the *lacI* gene) to the *lacO* operator sequence. The gene encoding the repressor is present in the bacterial chromosome and also in the pStabyGST1.2 plasmid to ensure high amount of repressor molecules. Consequently, in normal conditions, the T7 RNA polymerase is not or very weakly expressed. An additional repression of the *lac* promoter can be obtained using medium containing glucose (Grossman et al., 1998). The presence of glucose in the medium (especially in the stationary phase) induces the metabolic repression: the bacteria will first use glucose as a carbon source and will reduce the concentration of cyclic AMP, ensuring a better repression of the *lac* promoter (cyclic AMP stimulates the *lac* and *lacUV5* promoters). Moreover, Studier et al. (1990) have shown that a better regulation of the expression of the gene of interest is obtained by adding the *lacO* operator sequence between the T7 promoter and the beginning of the gene of interest. This sequence is present in the pStaby vectors. Consequently, the *lac* repressor will also repress the expression of the gene of interest.

Figure 1: The T7 expression system used in the SE1 strain



Adding isopropyl- β -D-thiogalactoside (IPTG) to the medium will induce the expression of (i) the T7 RNA polymerase and of (ii) the gene of interest by removing the *lac* repressor bound to the *lacO* sequence (figure 1).

A powerful feature of the T7 expression system is the ability to clone the gene of interest under conditions of extremely low or no transcriptional activity, that is, in the absence of the T7 RNA polymerase (as the CYS21 genetic background). The expression of the gene of interest is minimal in the absence of the T7 RNA polymerase because this gene is under the control of the T7 promoter which is only recognized by the T7 RNA polymerase and not by the *E. coli* RNA polymerase. If the target gene is cloned directly into the expression strain, even a low basal expression of the T7 RNA polymerase can interfere with growth and selection of the right construct. After the cloning step into a cloning strain lacking the T7 RNA polymerase (CYS21), the plasmid construct is transferred into the expression strain encoding the T7 RNA polymerase (SE1) to produce the protein of interest.

Overview of the stabilization system:

Higher plasmid stability= More proteins

Principle: In the StabyExpress™ system, the antidote gene (*ccdA*) is introduced in the plasmid DNA under the control of a constitutive promoter. On the other hand, the toxic gene (*ccdB*) is introduced in the chromosome of the bacteria (cf. fig. below). Expression of the poison gene is under the control of a promoter strongly repressed in the presence of the plasmid. When the plasmid is lost, the antidote is degraded and the production of the toxin is induced, causing cell death.

Experimental outline: Easy 4 steps procedure.

1. Cloning of your gene of interest in the pStabyGST vector
- ↓
2. Transformation into the CYS21 *E. coli* cells and selection of the desired construction
- ↓
3. Transformation of your plasmid DNA into the SE1 *E. coli* competent cells
- ↓
4. Expression of your gene of interest without antibiotic

Step 1: Cloning of your gene of interest in the pStabyGST1.2 vector:

Many strategies can be used for cloning your gene of interest (goi) into the pStabyGST1.2 vector. The most convenient strategy is to use restriction enzymes: the single-cutter enzymes from the multiple cloning sites are indicated on the map (see below). Use the buffer and incubation conditions provided by the restriction enzyme manufacturer. After restriction, the DNA fragment encoding the gene of interest is inserted by ligation using compatible ends.

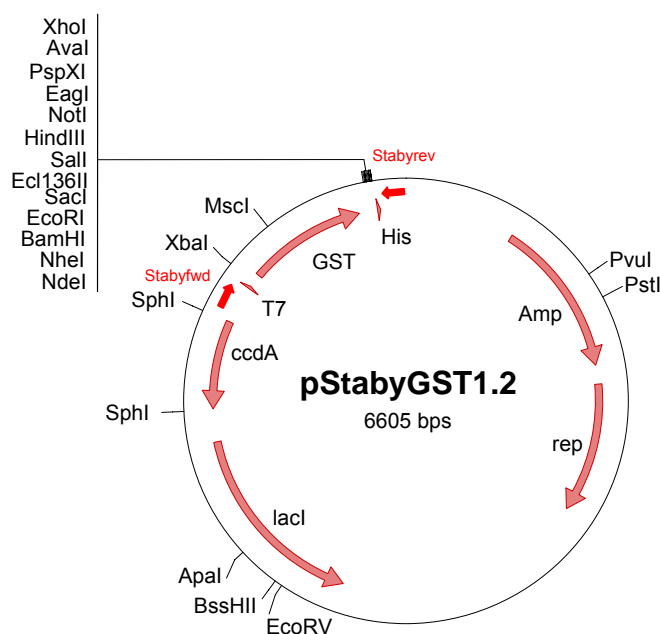


Figure 3: Restriction map of the pStabyGST1.2 vector

- Features:
- Staby forward primer: 5566-5584
 - T7 promoter: 5606-5622
 - GST tag: 5695-6348
 - Enterokinase cleavage site: 6349-6363
 - His: 6449-6466
 - Staby reverse primer: 6509-6491 (C)

The complete sequence of the vector is available on our website (www.delphigenetics.com)

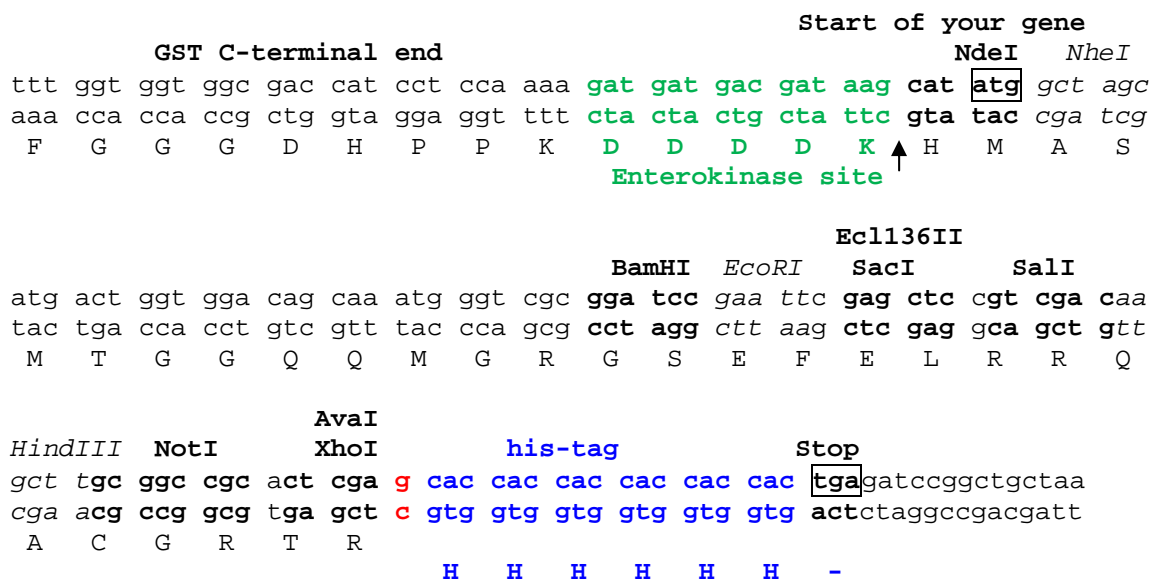
Remarks:

Take into account the reading frame of the GST gene (glutathione-S-transferase) to fuse your sequence to the GST tag at the N-terminal end of your protein (cf. figure below). This tag facilitates detection and purification of the target protein.

Note that an **Enterokinase site** is present in the pStabyGST1.2 vector at the C-terminal end of the GST tag to cleave the fused protein (removal of the GST tag). The cleavage position is indicated in the figure below (after the recognition site DDDDK).

The pStabyGST1.2 vector allows also the **fusion** of 6 histidine residues at the C-terminal end of the protein. If needed, the C-terminal fusion can be skipped by including a **stop codon** (TAA or TGA) at the end of your gene of interest. Please, note that the 6 histidine residues are not in the same frame than GST to avoid any fortuitous fusion (the 6 histidine codons are indicated in blue in the figure below). Consequently, you must take into account the reading frame of this His-tag if you want to use it.

Fig. 4. Sequence of the cloning region of pStabyGST1.2: (unique restriction sites are indicated in bold or italicized)

**Important:**

Delphi Genetics can help you with a software-based optimization of the nucleotide sequence of your gene-of-interest for a best protein production (please contact us at stabysoft@delphigenetics.com).

Step 2. Transformation into the CYS21 strain and selection of the desired construction:

Selection of the desired construction is performed in CYS21 *E. coli* cells lacking the T7 RNA polymerase gene (*). These cells contain the *ccdB* gene in their chromosome.

This enables:

- (i) High efficiency of transformation (the transformation efficiency of SE1 - derivative of BL21- is lower than that of CYS21),
- (ii) Stabilization of plasmids for high DNA production,
- (iii) Selection of the desired construction without expression of the gene of interest (goi).

(*)Remark:

It is not recommended to clone directly the goi into the expression host containing the T7 RNA polymerase gene: the T7 gene basal expression, and the resulting goi basal expression, would reduce the efficiency of recovery of the desired construction.

Protocol:

Two different types of StabyExpress™ kits are available: one containing electrocompetent cells, and the other type containing chemically-competent cells.

a) Transformation by electroporation:

- 1) Prepare LB plates containing 100µg/ml Ampicillin. Let the plates dry and then warm them at 37°C.
- 2) Set up your electroporator for bacterial transformation. Use the manufacturer's instructions. Classically, electroporation conditions are: 2,5 kV, 25 µF, and 200 Ohms.
- 3) For each cloning reaction, place one vial of the **CYS21** electrocompetent cells (pink cap) and one electroporation cuvette on ice. Allow the cells to thaw on ice for 5-10 minutes.
- 4) Add 1 or 2 µl of the ligation to the vial of the **CYS21** electrocompetent cells (pink cap). Stir gently to mix. Do not mix by pipetting up and down.

If you wish to use more than 2µl of the ligation mix, it is recommended to dialyze it against sterile water using a 0.025µm filter. Add the sterile water in a Petri dish and carefully place the filter on the water surface. Delicately, put the ligation mix on the filter. Wait 10min, pipet back the ligation mix and add the dialyzed solution to the electrocompetent cells.

- 5) Transfer all the content of the tube (cells+DNA) to the pre-chilled electroporation cuvette.
- 6) Electroporate the cells according to the manufacturer's instructions.
- 7) Quickly add 500µl of the regeneration medium (white cap) at room temperature and mix well.

- 8) Spread immediately 20 to 150µl on the pre-warmed plates containing ampicillin.
- 9) Incubate the plates overnight at 37°C.
- 10) Pick about 10 colonies and culture them overnight in 10ml of LB medium with or without ampicillin (100µg/ml).

Note: The stabilization is now effective; the ccdB gene is activated. Consequently, the plasmid is stabilized in the CYS21 strain and no antibiotic is needed to select bacteria containing the plasmid. However, the ampicilline resistance is still available. The stabilization system will insure high yield of plasmid DNA.

- 11) Extract plasmid DNA and analyze the constructions using your method of choice (restriction, sequencing, ...). Sequencing primers (**Staby reverse** and **Staby forward** primers) are included in the kit (0.1 µg/µl). The complete sequence of the pStabyGST1.2 vector is available on our website: <http://www.delphigenetics.com>.
- 12) Choose one of the clones containing the desired construct. Mix well 800µl of the culture with 800µl of sterile glycerol and transfer to a cryo-vial. Store at -80°C.

b) Transformation using chemically competent cells:

1. Prepare LB plates containing 100 µg/ml Ampicillin. Let the plates dry and then warm them up at 37°C.
2. Set a water bath or a heating-bloc to 42°C
3. Thaw (bring to room temperature) one vial of regeneration medium (white cap) per cloning reaction.
4. For each cloning reaction, place one vial of the **CYS21** chemically-competent cells (self-standing tube with pink cap) on ice. Allow the cells to thaw on ice for 5-10 minutes.
5. Add 5 µl of the ligation product to one vial of the **CYS21** chemically competent cells (self-standing tube with pink cap). Stir gently to mix. Do not mix by pipetting up and down.
6. Incubate on ice for 30 minutes.
7. Heat-shock the bacteria by placing the vial at 42°C for 30 seconds without shaking.
8. Immediately transfer the tubes to ice.
9. Add 250µl of room-temperature regeneration medium (white cap) and mix well.
10. Spread immediately 10, 20 and 100µl of the product (from step 9) on different pre-warmed plates.

If you wish to have more clones, incubate the product (from step 9) at 37°C for one hour for regeneration of the bacteria before spreading of 10, 20 and 100µl on different pre-warmed plates.

11. Incubate the plates overnight at 37°C.
12. Pick about 10 colonies and culture them overnight in 10ml of LB medium with or without ampicillin (100µg/ml).

Note: The stabilization is now effective; the ccdB gene is activated. Consequently, the plasmid is stabilized in the CYS21 strain and no antibiotic is needed to select bacteria containing the plasmid. However,

the ampicilline resistance is still available. The stabilization system will insure high yield of plasmid DNA.

13. Extract plasmid DNA and analyze the constructions using your method of choice (restriction, sequencing, ...). Sequencing primers (**Staby reverse** and **Staby forward** primers) are included in the kit (0.1 µg/µl). The complete sequence of the pStabyGST1.2 vector is available on our website: <http://www.delphigenetics.com>
14. Choose one of the clones containing the desired construct. Mix well 800µl of the culture with 800µl of sterile glycerol and transfer to a cryo-vial. Store at -80°C.

Step 3. Transformation in the expression host:

Positive control: The pStabyGST1.2 vector can be used without gene of interest as a positive control of your experiment. To do so, transform it according to the protocol below.

a) Transformation by electroporation:

- 1) Prepare LB plates containing 100µg/ml Ampicillin. Let the plates dry and then warm them at 37°C.
Note: Addition of 1% glucose (from a sterile filtered 20% stock solution) in the plates can be useful to better repress the promoter and to avoid basal expression.
- 2) Set up your electroporator for bacterial transformation. Use the manufacturer's instructions. Classically, electroporation conditions are: 2,5 kV, 25 µF, and 200 Ohms.
- 3) For each transformation, place one vial of the **SE1** electrocompetent cells (blue cap) and one electroporation cuvette on ice. Allow the cells to thaw on ice for 5-10 minutes.
Note: to use the pStabyGST1.2 vector without gene of interest as a control, do not forget to add an additional vial of SE1 and an additional cuvette.
- 4) Add 1µl of the selected plasmid DNA (steps 11 and 12 above or pStabyGST1.2 without gene of interest) to the **SE1** cells and mix gently.
- 5) Transfer all the content of the tube (cells+DNA) to the pre-chilled electroporation cuvette.
- 6) Electroporate the cells according to the manufacturer's instructions.
- 7) Quickly add 500µl of the regeneration medium (white cap) at room temperature and mix well.
- 8) Spread immediately 20 to 150µl on the pre-warmed LB plates.
- 9) Incubate the plates overnight at 37°C.
- 10) Optional: Pick about 5 colonies and culture them overnight in 10ml of LB medium.

Note: The plasmid is now stabilized in the SE1 strain using the StabyExpress™ system, no antibiotic is needed to select bacteria containing the plasmid. However, the ampicilline resistance

gene is still available. Addition of 1% glucose (from a sterile filtered 20% stock solution) in the medium can be useful to better repress the promoter and to avoid basal expression.

- 11) Optional: Extract plasmid DNA and analyze the constructions using your method of choice (restriction, sequencing, ...). Sequencing primers are included in the kit (0.1 µg/µl). The complete sequence of the pStabyGST1.2 vector is available on our website: <http://www.delphigenetics.com>
- 12) Optional: Select one of the clones containing the desired construction. Mix well 800µl of the culture with 800µl of sterile glycerol and transfer to a cryovial. Store at -80°C.

b) Transformation using chemically competent cells:

1. Prepare LB plates containing 100 µg/ml Ampicillin. Let the plates dry and then warm them up at 37°C.

Note: Addition of 1% glucose (from a sterile filtered 20% stock solution) in the plates can be useful to better repress the promoter and to avoid undesirable expression.

2. Set a water bath or a heating-bloc to 42°C
3. Thaw (bring to room temperature) one vial of regeneration medium (white cap) per cloning reaction.
4. For each transformation, place one vial of the **SE1** chemically-competent cells (self-standing tube with blue cap) on ice. Allow the cells to thaw on ice for 5-10 minutes.

Note: to use the pStabyGST1.2 vector without gene of interest as a positive control, do not forget to thaw an additional vial of SE1.

5. Add 1µl or 2µl of the selected plasmid DNA (steps 11 and 12 above or the pStabyGST1.2 without gene of interest as a control) to one vial of the **SE1** chemically competent cells (self-standing tube with blue cap). Stir gently to mix. Do not mix by pipetting up and down.
6. Incubate on ice for 30 minutes.
7. Heat-shock the bacteria by placing the vial at 42°C for 30 seconds without shaking.
8. Immediately transfer the tubes to ice.
9. Add 250µl of room-temperature regeneration medium (white cap) and mix well.
10. Spread immediately 10, 20 and 100µl of the product (from step 9) on different pre-warmed plates.
11. Incubate the plates overnight at 37°C.
12. Optional: Pick about 5 colonies and culture them overnight in 10ml of LB medium.

Note: The plasmid is now stabilized in the SE1 strain using the StabyExpress™ system, no antibiotic is needed to select bacteria containing the plasmid. However, the ampicilline resistance gene is still available. Addition of 1% glucose (from a sterile filtered 20% stock solution) in the medium can be useful to better repress the promoter and to avoid basal expression.

13. Optional: Extract plasmid DNA and analyze the constructions using your method of choice (restriction, sequencing, ...). Sequencing primers are included

in the kit (0.1 µg/µl). The complete sequence of the pStabyGST1.2 vector is available on our website: <http://www.delphigenetics.com>

14. Optional: Select one of the clones containing the desired construction. Mix well 800µl of the culture with 800µl of sterile glycerol and transfer to a cryovial. Store at -80°C.

Step 4. Expression of your gene of interest:

The T7 RNA polymerase is under the control of the PlacUV5 promoter (Studier and Moffat, 1986; Studier *et al.*, 1990). Both the SE1 strain and the pStabyGST1.2 vector carry the *lacI* gene. LacI represses both the expression of the T7 RNA polymerase and the transcription of the gene of interest. Consequently, the expression of the T7 RNA polymerase is inducible by isopropyl-β-D-thiogalactoside (IPTG): addition of IPTG to the culture of the SE1 strain containing the pStabyGST1.2 plasmid will induce the expression of the T7 RNA polymerase which, in turn, will transcribe the gene of interest.

Expression can also be performed using **Staby™Switch medium** (auto-inducible medium without IPTG). Perform the small-scale expression following the manual instructions.

The small-scale protocols below will allow you to verify that the target protein is produced upon induction and to verify for the presence of detection tags in the target protein.

Experiment control: The SE1 bacteria containing the pStabyGST1.2 vector alone will allow you to test the expression of the GST protein alone (without his tag, 262 amino acids, molecular weight: 30.3kDa, presents as homodimer : ~60kDa).

Protocol for a small-scale expression using Staby™Switch auto-inducible medium:

1. Inoculate two containers containing the desired volume of pre-warmed *Staby™Switch* medium with a few microliters (1 or 2 µl / 10ml culture) from a glycerol stock of the SE1 strain containing your construction in the pStabyGST1.2 vector. Alternatively, inoculate containers with a single colony from a plate streaked with this strain.

For a 10ml culture volume, the use of 50ml tubes with conical bottom (28mm x 114mm) is ideal. The tubes can be maintained closed during all the whole expression experiment. For bigger culture volumes, use Erlenmeyer flasks with a capacity of 5 times the culture volume. For 96 well plates, use 1 single colony or 0.001 volume of a glycerol stock per well.

Antibiotics are not required but can be used.

2. Add 1% sterile glucose (from a sterile-filtered 20% stock solution) to one of the two containers. This culture will be used as a non-induced control and/or to prepare a glycerol stock.
3. Incubate the containers at 37°C for approximately 24 hours with shaking (200rpm max, rotary shaker, 2.54cm orbit).
*Note: (1) If your protein is unstable, add 1% lactose (from a sterile-filtered 20% stock solution) 2 hours before the end of the culture.
(2) It is essential to grow the bacteria to stationary phase for full induction. If you want to incubate your cultures at lower temperature (<37°C), it is necessary to adapt the incubation time. Continue incubation for several hours (8 to 10 hours) after saturation. The first time, it is recommended to take a sample every hour and to check the protein expression (red colour or SDS-PAGE analysis).*
4. After incubation, measure the Optical Density at 600nm for each culture. Transfer 1ml sample of each flask in a microcentrifuge tube. Centrifuge at maximum speed (13000 g) for 10 min (if possible at 4°C).

Protocol for a small-scale expression using IPTG:

- 1) Inoculate two Erlenmeyer flasks containing 10ml of LB medium with a few microliters from a glycerol stock. Alternatively, pick two single colonies from a plate streaked with the SE1 bacteria containing your construction; inoculate two flasks containing 10ml of LB medium.
- 2) Incubate with shaking at 37°C until OD₆₀₀ reaches 0.4-1 (the best range is between 0.6 and 0.8).
- 3) In one of the two flasks, add IPTG (100µl of a fresh 100mM stock solution) to reach a final concentration of 1mM. The other flask is used as a non-induced control. Continue incubation of both flasks for 2-3 hours.
- 4) Measure the Optical Density at 600nm for each culture. Transfer 1ml sample of each flask in a microcentrifuge tube.

Protein extraction under denaturing conditions

- 1) Transfer 1ml sample of each flask in a microcentrifuge tube. Centrifuge at maximum speed (13000 g) for 10 min (if possible at 4°C). Discard the supernatant, add 1ml H₂O and resuspend the bacteria. Add 50µl of cold 100% Trichloroacetic acid (TCA) (w/v) to each sample and vortex for a few seconds.

Note: The TCA precipitation allows the analysis of the total protein content of the cells. Other methods can be used to specifically analyze different fractions (soluble, insoluble, periplasm, ...) in order to identify the cellular localization of the target protein. For more information, please, check specialized literature or protocols (e.g., Sambrook et al., Ausubel et al.).

- 2) Place on ice for 10 min.
- 3) Centrifuge at maximum speed (13000 g) for 10 min (if possible at 4°C).
- 4) Remove carefully and discard the supernatant.

- 5) Wash the pellet with cold acetone (+4°C): add 500µl of acetone, vortex, and centrifuge for 5 min at maximum speed (if possible at 4°C).
- 6) Repeat steps 7 and 8
- 7) Remove carefully the supernatant. Air dry the final pellet: leave the tube opened on the bench or use vacuum drying.
- 8) Add (OD₆₀₀ x 200)µl of 1X sample buffer (2X sample buffer= 100mM DTT, 2% SDS, 80mM Tris-HCl, pH 6.8, 0.006% bromophenol blue, 15% glycerol). Vortex vigorously to resuspend the pellet.

Note: Taking into account the OD₆₀₀ allows comparison of Coomassie-stained band intensities between samples.

- 9) Heat the samples at 70°C-100°C (10min.) to resuspend and denature the proteins. The samples can be used directly for SDS-PAGE analysis or stored at -20°C.
- 10) Load 4 to 10 µl of each sample in a SDS-PAGE gel containing the appropriate concentration of polyacrylamide (according to the size of the overproduced protein). Add a molecular size marker.

Note: The sample volume that needs to be loaded will depend on the gel size, the expression level, and the extraction efficiency.

- 11) After migration, visualize the proteins with Coomassie-blue staining or continue the analysis with western blot.

Note: Western blot analysis is a more specific and sensitive method but needs protein-specific antibodies or fusion tag-specific antibodies. For more information, please, check specialized literature or protocols (e.g., Sambrook et al, Ausubel et al.).

About protein detection and purification using GST:

GST (Glutathione-S-Transferase) is commonly used for protein detection and purification. This protein is expressed in E. coli with full enzymatic activity. GST is a protein of 26 kDa with an isoelectric point pI = 5.0. The protein is soluble and presents as a **homodimer** in E. coli cytoplasm (molecular weight of the dimer: 58.5 kDa). In denaturing conditions, the dimer is resolved and a single band at ~26kDa is visible. However, if proteins were not completely denatured before loading on gel, the dimer is observed (band at ~58.5 kDa, even when using SDS-PAGE gels). It is thus necessary to take into account the presence of GST dimer when you analyse your gels when denaturing conditions were not used.

GST detection can be performed using SDS-PAGE analysis, enzymatic detection (using GST substrate 1-chloro-2,4 dinitrobenzene, CDNB) or Western blot. The Western blot is often the most powerful method. For more information about the use of anti-GST antibody, please, consult documentation of the manufacturer but do not forget to run controls (protein samples of E. coli that do and do not contain parental pStabyGST1.2 vector).

For protein purification using GST, several products exist and can be used with your fusion protein. One of the most useful is the glutathione-sepharose column: the glutathione is attached to sepharose and is complementary to the binding site of the GST binding site. One of the most important parameters affecting the binding of GST fusion proteins to glutathione-sepharose column is the flow rate. Since the

binding kinetics between glutathione and GST are relatively slow, it is necessary to keep the flow rate constant and low during sample application to achieve maximum binding capacity. Another point to consider is the protein solubility. GST is soluble and can help to solubilise your protein. However, if your fusion protein is insoluble, it is often easier to purify it using the His-tag of the vector: using (His)₆ fusion protein enables the use of denaturing agents during purification (and refolding after purification) or on-column refolding procedure. For more information about these procedures, please, consult documentation of the column manufacturer.

In most cases, functional tests can be performed using the fusion protein but if removal is necessary, the pStbayGST1.2 vector contains an enterokinase cleavage site at the C-terminal end of GST. This site is the specific cleavage site of the enterokinase protease. It allows the removal of GST after purification or detection without leaving extra amino acids. Take into account that after cleavage, the protease should be removed. Columns with specific affinity to enterokinase exist and could be combined to the purification column. For more information, please, contact the manufacturer.

Troubleshooting:

Please note that problems with cloning or expression efficiencies can result from the following parameters. Most of these problems can be fixed as explained below. However, due to intrinsic and specific properties of your gene or protein, the cloning or expression efficiencies may vary.

Problem	Solution
Only a few or no colonies are observed after transformation (ligation mix into the CYS21 bacteria or plasmid construct into the SE1 bacteria).	<ul style="list-style-type: none"> - Check the DNA concentration of your insert and the ligation conditions. - Check the quality of your insert (one single band must be visible after agarose gel electrophoresis of the purified DNA fragment). - Be sure that the DNA transformation was optimal. When using electrocompetent bacteria, check the electroporation conditions (see above). When using chemically competent bacteria: check the temperature of the water bath, incubate the transformation product during one hour at 37°C to allow regeneration of the bacteria before spreading. - Check your plates with another strain which is resistant to the ampicillin antibiotic. If no growth is observed, check your antibiotic solution. - Your cloned fragment could be toxic for the bacteria. Check the literature (if data are available). Add 1% glucose (from a sterile 20% stock solution) to the plates to better repress the promoter. - Test the transformation efficiency with the pStabyGST1.2 vector without gene of interest
No expression	<ul style="list-style-type: none"> - Check the gene sequence for mutations. - Check your expression conditions. - Check expression using the expression control (SE1 containing pStabyGST1.2 without gene of interest). - Check expression starting with a single colony or a glycerol stock (do not use pre-culture) - When using Staby™Switch medium, pre-warm the medium at room temperature or 37°C before inoculation. - Your protein might be unstable, try different induction conditions (lower temperature, Staby™Switch medium, longer induction time,...). - Use Cherry™Express or Cherry™Codon kits to follow your protein with your eyes (for more information, please consult our website: www.delphigenetics.com).

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Related Staby™ products and services:



The **StabyCloning™ kit** is designed for the rapid, precise and efficient DNA cloning of PCR products. The complete cloning procedure is performed in one hour (including plating), the background is basically nil (the bacteria containing vectors without insert are killed), the PCR product is oriented, the plasmid is stabilized, and the export of the insert to another vector is easily selected.



The **StabyExpress™ T7 kit** contains all the key elements for cloning of a gene-of-interest and its expression in *Escherichia coli*. The kit combines two technologies (T7 expression and plasmid stabilization) that allow high-yield protein expression and standardization of the production-protocol. Vectors with his-tag and/or GST-tag are available.



The **GetStaby™ kit** allows easy addition of Delphi-Genetics' stabilization technology into your favourite vector. The technology is compatible with any expression system. Using this technology, your vectors are perfectly stabilized even without antibiotics.



The **Staby™ Codon T7 kit** combines three technologies to ensure high-yield and standardized expression of eukaryote proteins in *Escherichia coli*. These technologies are (i) T7-controlled expression, (ii) plasmid stabilization, and (iii) codon-usage adaptation of *E. coli* for the efficient expression of proteins that contain rare codons.



The **Cherry™ Express kit** allows direct visualization (by eye!) of your protein of interest during protein production in *E. coli* and protein purification. Special requirements or reagents are not needed. It is also possible to quantify the protein concentration at any step by spectral measurement. The Cherry™ Express kit combines multiple advantages: protein visualization, T7 expression, plasmid stabilization and codon-usage adaptation.



The **Staby™ Switch** medium is an auto-inducible medium (ready-to-use) designed for high-level protein expression using Staby™ products or any other IPTG-inducible bacterial expression system. Using Staby™ Switch medium, protein expression is automatically induced when high cell density is reached. Thus, it is neither necessary to add IPTG nor to monitor optical density during bacterial growth.



Staby™ Soft was specifically designed by Delphi Genetics to support the users of the Staby™ Operating System. This software package can perform customized gene-of-interest analysis to choose the most adapted kit and to optimize protein production.

For more information, please, consult www.delphigenetics.com

Staby™ products ordering information:

StabyExpress™		
GE-SET7-0505	pStaby1.2 expression kit, electro-competent	5 reactions
GE-SET7-0707	pStaby1.2 expression kit, chemically-competent	5 reactions
GE-SET7-1010	pStaby1.2 expression kit, electro-competent	10 reactions
GE-SET7-1212	pStaby1.2 expression kit, chemically-competent	10 reactions
GE-SET7-2020	pStaby1.2 expression kit, electro-competent	20 reactions
GE-SET7-2222	pStaby1.2 expression kit, chemically-competent	20 reactions
GE-SEGST-0505	pStabyGST1.2 expression kit, electro-competent	5 reactions
GE-SEGST-0707	pStabyGST1.2 expression kit, chemically-competent	5 reactions
GE-SEGST-1010	pStabyGST1.2 expression kit, electro-competent	10 reactions
GE-SEGST-1212	pStabyGST1.2 expression kit, chemically-competent	10 reactions
GE-SET7-1111	10 x 50µl CYS21 and 10x50µl SE1 bacteria, electro-competent	10 reactions
GE-SET7-1313	10 x 100µl CYS21 and 10x100µl SE1 bacteria, chimio-competent	10 reactions
GE-SET7-0020	Set of 20 expression bacteria (SE1), electro, 50µl/tube	20 reactions
GE-SET7-0022	Set of 20 expression bacteria (SE1), chimio, 100µl/tube	20 reactions
GetStaby™		
GE-GSA1-10	GetStaby kit, electro-competent cells	10 reactions
GE-GSA1-12	GetStaby kit, chemically-competent cells	10 reactions
StabyCloning™		
GE-STC1-10	StabyCloning kit, electro-competent cells	10 reactions
GE-STC1-12	StabyCloning kit, chemically-competent cells	10 reactions
GE-STC1-20	StabyCloning kit, electro-competent cells	20 reactions
GE-STC1-22	StabyCloning kit, chemically-competent cells	20 reactions
GE-STCB-20	Set of 20 cloning bacteria (CYS21) electro (50µl/tube)	20 reactions
GE-STCB-22	Set of 20 cloning bacteria (CYS21) chimio (100µl/tube)	20 reactions
Staby™Codon		
GE-SCT7-0505	pSCodon1.3 expression kit, electro-competent	5 reactions
GE-SCT7-0707	pSCodon1.3 expression kit, chimio-competent	5 reactions
GE-SCT7-1010	pSCodon1.3 expression kit, electro-competent	10 reactions
GE-SCT7-1212	pSCodon1.3 expression kit, chimio-competent	10 reactions
GE-SCGST-0505	pSCodonGST1.2 expression kit, electro-competent	5 reactions
GE-SCGST-0707	pSCodonGST1.2 expression kit, chimio-competent	5 reactions
GE-SCGST-1010	pSCodonGST1.2 expression kit, electro-competent	10 reactions
GE-SCGST-1212	pSCodonGST1.2 expression kit, chimio-competent	10 reactions
Staby™Switch		
GE-AIME-04	Auto-induction medium	2L
Cherry™Express		
GE-CET7-05	CherryExpress T7 expression kit, N-terminal tag, electro	5 reactions
GE-CET7-06	CherryExpress T7 expression kit, C-terminal tag, electro	5 reactions
GE-CET7-07	CherryExpress T7 expression kit, N-terminal tag, chimio	5 reactions
GE-CET7-08	CherryExpress T7 expression kit, C-terminal tag, chimio	5 reactions
GE-CET7-10	CherryExpress T7 expression kit, N-terminal tag, electro	10 reactions
GE-CET7-11	CherryExpress T7 expression kit, C-terminal tag, electro	10 reactions
GE-CET7-12	CherryExpress T7 expression kit, N-terminal tag, chimio	10 reactions
GE-CET7-13	CherryExpress T7 expression kit, C-terminal tag, chimio	10 reactions
Cherry™Codon		
GE-CCT7-05	CherryCodon T7 expression kit, N-terminal tag, electro	5 reactions
GE-CCT7-06	CherryCodon T7 expression kit, C-terminal tag, electro	5 reactions
GE-CCT7-07	CherryCodon T7 expression kit, N-terminal tag, chimio	5 reactions
GE-CCT7-08	CherryCodon T7 expression kit, C-terminal tag, chimio	5 reactions
GE-CCT7-10	CherryCodon T7 expression kit, N-terminal tag, electro	10 reactions
GE-CCT7-11	CherryCodon T7 expression kit, C-terminal tag, electro	10 reactions
GE-CCT7-12	CherryCodon T7 expression kit, N-terminal tag, chimio	10 reactions
GE-CCT7-13	CherryCodon T7 expression kit, C-terminal tag, chimio	10 reactions

**ACADEMIC AND NON-PROFIT LABORATORY ASSURANCE LETTER REGARDING
THE USE OF THE T7 EXPRESSION SYSTEM**

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